UDC: 616.314-084-053.2:612.392.69:546.15

Analysis of biocoenosis of oral cavity in children with different degrees of caries actuvity, who permanently residing in conditions of biogeodeficit of fluorine and iodine.

O.V. Klitynska

Department of Dentistry of Childhood Age, State Higher Educational Establishment "Uzhgorod National University", Uzhgorod, Ukraine

Summary: Microbial scenery of the oral cavity plays an important role in the formation and progression of dental disease that is particularly revealing in preschool and early school age. Research microbiota of the oral cavity, species identification and quantitative composition marker is the origin and progression of diseases of the oral cavity as caries, inflammatory and degenerative inflammatory diseases periodontal tissues and lesions of the oral mucosa.. **Key words:** oral cavity, careis,

biogeodeficit, microbiota.

Actuality. Microbial scenery of the oral cavity plays an important role in the formation and progression of dental disease that is particularly revealing in preschool and early school age. According to different authors in the oral cavity occurs more than 100 species of microorganisms, and 1 ml saliva

contains more than 108 microbial cells [6-9]. prevalence of certain kinds The microorganisms creates conditions for the formation and rapid progression of dental disease [1,5,10,11]. Evaluation of microbial scenery plays an important role in the choice of medical tactics and as a result improve the effectiveness of treatment [2-4,12-14].For setting the microbial scenery oral saliva sampling was carried out and microbial soft plaque in the cervical area. Hung held the following nutrient medium: - meat peptonnyy agar to determine TMC (total microbial count) - agar selective for streptococci - selective agar for Streptococcus mitis and S. salivarius; semi-selective medium for the isolation of streptococci - semi environment for the allocation of bifidobacteria (environment Blauroka) - Endo environment to check for gram-negative microbiota - bismuth-sulphite agar for the isolation of Salmonella and Shigella - enterokokovyy agar for the cultivation of species of Entecococcus spp.; laktobak-agar (MRS - Man -Rogosa-Sharpe agar) for the cultivation of lactobacilli in microaerophilic conditions bifidum, Bifidobacterium Agar for cultivation.

For the detection and identification using Gram smears production and further their microscoping, routine biochemical tests. All relevant strains isolated during the experiment strains of microorganisms will be saved.

The study was conducted in 146 children with different degrees of caries activity, aged 3-8 years, who permanently live in conditions of biochemical deficits of different ethnic groups. A survey of children was conducted after informed consent of parents or guardians of LLC "Dental clinic" Dental Faculty SHEI "Uzhhorod National University". Microbiological and immunological studies were conducted in microbiological laboratories Transcarpathian

V.II (4) / 2015

Ukraine Branch of microbiologists name S.M.Vynohradskoho - TUBM.

Results and discussion. The results of determining microbiota in saliva of inspected children, Roma ethnic group, mixed ethnic groups and the control group presented in Figure 1.

Average value of total microbial number of children in the control group was 4 \times 105, not significantly different rates in children of the Roma ethnic group 6 \times 105 (p> 0.05)., And the children of mixed ethnic groups was significantly higher and was 6 \times 109 (p <0.05).

Among the species belonging were significant differences among children of different groups. So, children of mixed ethnic groups were representatives of normal oral microflora and opportunistic and pathogenic microorganisms.

Normal microflora presented oral Streptococcus mitis, S. salivarius, Lactobacillus salivarius, Bifidobacterium spp. - normally found in the mouth, including saliva, plaque, in the mucous membranes of the mouth. Number of microbiota in children of the main group was small: Streptococcus mitis - 1×104 CFU; Streptococcus salivarius - 2×104 cfu; Bifidobacterium spp - 1×104 CFU; Lactobacillus salivarius - 8×102 CFU; Lactobacillus acidophilius - 8×101 CFU.

Streptococcus sobrinus - typical caries forming species with Streptococcus mutans to form a group of similar species Streptococcus mutans biochemical properties and effects on the surface of the tooth enamel and tooth tissue destruction. owed by 100% of children of the main group, the average values were Streptococcus sobrinus - 4×104 CFU.

Other species, including representatives of gram-negative microbiota and species of the genus Enterococcus spp. and Staphylococcus aureus normally have to be present among the normal microbiota; Data representatives (Escherichia coli - 6×106 CFU; Proteus vulgaris - 8×108 CFU; Citrobacter freundii - 2×107 cfu; Pseudomonas aeruginosa - 1×109 CFU; Enterococcus faecalis - 1×105 CFU; Enterococcus faecium - 1×103 CFU) is normobiota and opportunistic bacteria by the gastrointestinal tract. A Staphylococcus aureus is the causative agent of many infections and diseases causing opportunistic and nosocomial infections in people with lowered immunity sown amount of 1×103 CFU.

This oral microbial scenery corresponds to the third degree of dysbiosis V.V.Hazanovoyu for all children of the main group, regardless of the degree of activity of dental caries, and is characterized by the detection of pathogenic monoculture with a sharp decrease in the number or total absence of representatives of normal (physiological) microflora.

The children of the Roma ethnic group frequency of sowing microflora significantly different. So, the average total microbial number was 6×105 . Flora represented Streptococcus mitis - 9×103 CFU; Streptococcus salivarius - 1×104 CFU; Bifidobacterium spp - 2×107 cfu; Lactobacillus salivarius and Lactobacillus acidophilius available.

Among the opportunistic pathogenic Enterococcus faecalis - 1×105 CFU. All members of the Roma ethnic group were allocated Streptococcus mutans in quantities of 1×104 CFU and Streptococcus sobrinus not sown. Accordingly, in this group of patients diagnosed subcompensated form of dysbiosis, which is characterized by the presence of one or two kinds of opportunistic microorganisms against the background of a reduction in titer lactobacilli.

In the control group inspected flora represented Streptococcus mitis - 9×103

CFU; Streptococcus salivarius - 1×106 CFU; Bifidobacterium spp, Lactobacillus salivarius and Lactobacillus acidophilius available. opportunistic presented Enterococcus faecalis -9 \times 103 cfu. The average values of total microbial count was 4 \times 105.

Ac	cid-st	reptokky	are	e	Streptococcus			
sobrinus	in	number	9	×	103	CFU;		

	Table 1												
		Streptococcus mitis	Streptococcus sobrinus	Psevdomonas aeruginosa	Enterococcus faecalis	Enterococcus faecium	Proteus vulgaris	Lactobacillus salivarius	Lactobacillus acidophilus	Staphylococcus aureus	Bifidobacterium	Citrobacter freundii	Escherichia coli
Streptococcus mitis	r	1	-0.03632	-0.07963	-0.02141	-0.04393	0.02341	0.21537	0.46036	-0.07302	0.77805	-0.07387	-0.07985
Streptococcus mitis	σ		8.36E-01	6.49E-01	9.03E-01	8.05E-01	8.94E-01	0.21404	5.39E-03	6.77E-01	3.78E-08	6.73E-01	0.6484
Streptococcus sobrinus	r	-0.03632	1	-0.04681	-0.07599	0.99134	-0.03547	-0.04883	-0.00636	0.23332	-0.02791	-0.02339	-0.0419
Streptococcus sobrinus	σ	8.36E-01		7.89E-01	6.64E-01	0	8.40E-01	0.78057	9.71E-01	0.17737	8.74E-01	8.94E-01	0.8111
Psevdomonas aeruginosa	r	-0.07963	-0.04681	1	-0.03722	-0.03155	-0.06418	0.92528	-0.05538	-0.12008	-0.04147	-0.0485	-0.03102
Psevdomonas aeruginosa	σ	6.49E-01	7.89E-01		8.32E-01	8.59E-01	7.14E-01	2E-15	7.52E-01	4.92E-01	8.13E-01	7.82E-01	8.60E-01
Enterococcus faecalis	r	-0.02141	-0.07599	-0.03722	1	-0.0605	-0.1068	-0.04136	-0.05318	0.22091	-0.02369	-0.03769	0.97643
Enterococcus faecalis	σ	9.03E-01	6.64E-01	8.32E-01		0.73396	5.41E-01	0.81351	7.62E-01	2.02E-01	8.93E-01	8.30E-01	0
Enterococcus faecium	r	-0.04393	0.99134	-0.03155	-0.0605	1	-0.03461	-0.04216	-0.05415	0.2357	-0.04585	-0.03992	-0.0303
Enterococcus faecium	σ	8.05E-01	0	8.59E-01	0.73396		8.46E-01	0.81288	0.76099	1.80E-01	7.97E-01	0.82264	8.65E-01
Proteus vulgaris	r	0.02341	-0.03547	-0.06418	-0.1068	-0.03461	1	-0.08462	-0.10864	0.36465	-0.08301	0.15265	-0.06085
Proteus vulgaris	σ	8.94E-01	8.40E-01	7.14E-01	5.41E-01	8.46E-01		6.29E-01	5.34E-01	3.13E-02	6.35E-01	3.81E-01	0.7284
Lactobacillus salivarius	r	0.21537	-0.04883	0.92528	-0.04136	-0.04216	-0.08462	1	0.186	-0.17998	0.33466	-0.0766	-0.03123
Lactobacillus salivarius	σ	0.21404	0.78057	2E-15	0.81351	0.81288	6.29E-01		0.28471	0.30085	4.94E-02	6.62E-01	0.85867
Lactobacillus acidophilus	r	0.46036	-0.00636	-0.05538	-0.05318	-0.05415	-0.10864	0.186	1	-0.23537	0.5945	-0.09864	-0.05251
Lactobacillus acidophilus	σ	5.39E-03	9.71E-01	7.52E-01	7.62E-01	0.76099	5.34E-01	0.28471		1.73E-01	1.66E-04	5.73E-01	7.65E-01
Staphylococcus aureus	r	-0.07302	0.23332	-0.12008	0.22091	0.2357	0.36465	-0.17998	-0.23537	1	-0.18186	0.41907	0.2231
Staphylococcus aureus	σ	6.77E-01	0.17737	4.92E-01	2.02E-01	1.80E-01	3.13E-02	0.30085	1.73E-01		2.96E-01	1.22E-02	1.98E-01
Bifidobacterium	r	0.77805	-0.02791	-0.04147	-0.02369	-0.04585	-0.08301	0.33466	0.5945	-0.18186	1	-0.07844	-0.03421
Bifidobacterium	σ	3.78E-08	8.74E-01	8.13E-01	8.93E-01	7.97E-01	6.35E-01	4.94E-02	1.66E-04	2.96E-01		6.54E-01	8.45E-01
Citrobacter freundii	r	-0.07387	-0.02339	-0.0485	-0.03769	-0.03992	0.15265	-0.0766	-0.09864	0.41907	-0.07844	1	-0.03126
Citrobacter freundii	σ	6.73E-01	8.94E-01	7.82E-01	8.30E-01	0.82264	3.81E-01	6.62E-01	5.73E-01	1.22E-02	6.54E-01		0.85853
Escherichia coli	r	-0.07985	-0.0419	-0.03102	0.97643	-0.0303	-0.06085	-0.03123	-0.05251	0.2231	-0.03421	-0.03126	1
Escherichia coli	σ	0.6484	0.8111	8.60E-01	0	8.65E-01	0.7284	0.85867	7.65E-01	1.98E-01	8.45E-01	0.85853	

Microbiota. Internal correlation

Pearson correlation coefficients "r" and the level of reliability " σ " for different pairs of microorganisms

All patients with decompensated caries had established the presence of oral

dysbiosis 3-4 degrees. These figures correspond dysbiotic change of 1 degree.

INTERMEDICAL JOURNAL

V.II (4) / 2015



Figure1.Determination of various types of microorganisms and secretory immunoglobulin A in saliva inspected depending on the ethnic group.

V.II (4) / 2015

INTERMEDICAL JOURNAL

Were following the bacteria: Streptococcus sobrinus - 100%; Streptococcus mitis - in 88.3%; Streptococcus salivarius - at 82.4%; Escherichia coli - 5.9%; Proteus vulgaris - at 47.1%; Citrobacter freundii - at 53.1%; Pseudomonas aeruginosa in 47.1%; Lactobacillus salivarius - at 11, 8% of cases (there are doubts about the plant species, 97% accuracy, using biochemical test Erba-Lachema, needs clarification on MALDI); Staphylococcus aureus at 76.5%: _ Enterococcus faecalis 88.3%; _ in Enterococcus faecium _ 5.9%: Bifidobacterium spp.- in 53.0% (giving

Streptococcus sobrinus - typical cavities forming species with Streptococcus mutans to form a group of similar species Streptococcus mutans biochemical properties and effects on the surface of the tooth enamel and tooth tissue destruction. One of the most important factors of virulence is its acid stability. In the analysis of the mechanism kyslotostiykosti streptokoiv and adaptation to acidic environment found the that Streptococcus sobrinus can continue its growth and reproduction in acidity pH 5.0, in such circumstances it continues to ferment carbohydrates through glycolysis and continues to acidification of the environment around them, with specific enzyme System sobrinus (including Streptococcus fosfotransferazna system) in a state of higher activity at pH 5.0 than in a neutral

Conclusions. Research microbiota of the oral cavity, species identification and quantitative composition marker is the origin and progression of diseases of the oral cavity as caries, inflammatory and degenerative inflammatory diseases periodontal tissues and lesions of the oral mucosa. In particular, Streptococcus sobrinus, which was allocated 100% of the surveyed children aged 4-6 years questionable results (65%) using biochemical anaerotestu Erba-Lachema, needs clarification on MALDI).

Streptococcus mitis, S. salivarius, Lactobacillus salivarius, Bifidobacterium spp. - Normally found in the oral cavity, including saliva, plaque, in the mucous membranes of the oral cavity.

Streptococcus mitis, S. salivarius, Lactobacillus salivarius, Bifidobacterium spp. - Normally found in the oral cavity, including saliva, plaque, in the mucous membranes of the oral cavity.

environment, creating conditions for the emergence of enamel demineralization, especially deciduous teeth in children [7, 12-14].

Other species, including representatives of gram-negative microbiota and species of the genus Enterococcus spp. and Staphylococcus aureus normally have to be present among the normal microbiota; Representatives of data coli. (Escherichia Proteus vulgaris, Citrobacter freundii, Pseudomonas faecalis. aeruginosa, Enterococcus E. faecium) is normobiotoyu and opportunistic bacteria by the gastrointestinal tract. A Staphylococcus aureus is the causative agent of many infections and diseases causing opportunistic and nosocomial infections in people with reduced immunity.

residing near biogeochemical fluorine and iodine deficiency, considered the most acidophilus bacteria that persist in the mouth of man because of bacteria colonies in rN5, 0 still active synthetic processes, they lack the shock and stress protein that has been proven to hemostatniy culture, DNA, RNA analysis and Western blotting.

Their timely identification and efficient removal of toxic effects by reducing their

number will reduce the risk of diseases of the oral cavity and create conditions for proper

formation of all the structures of the oral cavity in children.

REFERENCES

1. Захаров А.А. Анализ микрофлоры ротовой полости обследованных людей с различными заболеваниями / А.А. Захаров, Н.А. Ильина // Успехи современного естествознания. - 2007. - № 12. - С. 353 - 355.

2. Кузнецов, Е.В. Микробная флора полости рта и её роль в развитии; патологических процессов / Е.В.Кузнецов, В.Н.Царёв. — М.: МЕДпресс-информ, 2003. – 240 с.

3. Леус П.А. Биофильм на поверхности зуба и кариес / П.А. Леус. М.: Издательский Дом «STBOOK», 2008.- 88с.

4. Тец В.В. Роль микрофлоры полости рта в развитии заболеваний человека / В.В. Тец // Стоматология, 2008. №3. - С.76-80.

5. Al-Asqah M. Is the presence of Helicobacter pylori in the dental plaque of patients with chronic periodontitis a risk factor for gastric infection?/ M. Al-Asqah, N. Al-Hamoudi, W. K. Al-Hamoudi // Can J Gastroenterol. – 2009. – Vol. 23(3). – P.177–179

6. Belstrum D. Bacterial profiles of saliva in relation to diet, lifestyle factors, and socioeconomic status/ D. Belstrum, P. Holmstrup, C. H. Nielsen et al. // J Clin Periodontol. -2014 - Vol.41(2) - P. 104 - 112.

7. Haffajee A.D. Microbial complexes in supragingival plaque / S.S. Socransky, A.D. Haffajee, M.R. Patel, X. //Song Oral Microbiol Immunol. - 2008. - №23. - P.196–205.

8. Jia C.L. Effect of dental plaque control on infection of Helicobacter pylori in gastric mucosa / C.L. Jia, G.S. Jiang, C.H. Li, C.R. Li // Tex Dent J. 2012 Vol.129(10). P.1069-1073.

9. <u>Kaakoush N.O. Campylobacter concisus - A new player in intestinal disease / N.O.</u> Kaakoush, H.M. Mitchell // Front Cell Infect Microbiol. – 2012. - № 2. _ P.2-4.

10. Ly M. Altered Oral Viral Ecology in Association with Periodontal Disease / M. Ly, S. R. Abeles, T. K. Boehm et al. // MBio. - 2014. – Vol. 5, № 3. – P. 1133-1147.

Mahajan A. Interspecies Communication and Periodontal Disease/ A. Mahajan,
B. Singh, D. Kashyap et al. // The Scientific World Journal. – 2013. - Vol.2013. – P. 765434

12. Said H.S. Dysbiosis of Salivary Microbiota in Inflammatory Bowel Disease and Its Association With Oral Immunological Biomarkers/ H. S. Said , W. Suda., S. Nakagome et al. // DNA RESEARCH. – 2014. - N_{2} 21. – P.15–25.

13. <u>Salazar C.R. Association between selected oral pathogens and gastric precancerous</u> <u>lesions/ C.R. Salazar, J.Sun, Y. Li et al. // PLoS One.</u> – 2013. № 8(1). – P.e51604.

14. Socransky S.S. Microbial complexes in subgingival plaque / S.S. Socransky, A.D. Haffajee, M.A. Cugini et al.// J Clin Periodontol. – 1998. - №25. – P.134–144.